

糖原 PAS 染色液

产品简介：

糖原染色是病理学中常规的染色方法之一，McManus 在 1946 年最先使用高碘酸-雪夫技术显示黏蛋白，该法常用来显示糖原和其他多糖，该染色不仅能够显示糖原，还能显示中性黏液性物质和某些酸性物质以及软骨、垂体、霉菌、真菌、色素、淀粉样物质、基底膜等；过碘酸(又称高碘酸)是一种强氧化剂，它能氧化糖类及有关物质中的 1, 2-乙二醇基，使之变为二醛，醛与 Schiff 试剂能结合成一种品红化合物，产生紫红色，由于高碘酸还可氧化细胞内其他物质，使用时应注意选择好高碘酸浓度和氧化时间，使氧化控制在既能把乙二醇基氧化成醛基又不至于过氧化，这是很关键的步骤。

Leagene 糖原 PAS 染色液特点是采用 Leagene 特有配方技术，大大增强了染色效果；性能稳定，特异性强；操作简捷，仅需 1h 左右。该试剂仅用于科研领域，不适用于临床诊断或其他用途。

产品组成：

名称 \ 编号	DG0005 4×50ml	DG0005 4×100ml	DG0005 4×500ml	Storage
试剂(A): 过碘酸溶液	50ml	100ml	500ml	4℃ 避光
试剂(B): Schiff Reagent	50ml	100ml	500ml	4℃ 避光
试剂(C): Leagene 苏木素染色液	50ml	100ml	500ml	RT
试剂(D): 酸性乙醇分化液	50ml	100ml	500ml	RT
使用说明书	1 份			

自备材料：

- 1、10%福尔马林固定液
- 2、蒸馏水、系列乙醇、二甲苯或环保脱蜡透明液、中性树胶

操作步骤(仅供参考)：

- 1、常规固定，常采用 10%福尔马林固定液，常规脱水包埋。
- 2、石蜡切片二甲苯或 Leagene 脱蜡透明液脱蜡入蒸馏水；冰冻切片直接入蒸馏水。
- 3、自来水冲洗 2~3min，再用蒸馏水浸洗 2 次。
- 4、入过碘酸溶液室温放置 5~8min，一般不宜超过 10min。
- 5、自来水冲洗 1 次，再用蒸馏水浸洗 2 次。
- 6、入 Schiff Reagent 置于室温阴暗处浸染 10~20min，自来水冲洗 10min。

苏木素染色液，染细胞核 1 ~ 2min，酸性乙醇分化液分化 2 ~ 5s。

- 8、自来水冲洗 10 ~ 15min，更换蒸馏水清洗，使其返蓝。
- 9、逐级常规乙醇脱水，二甲苯或 Leagene 脱蜡透明液透明，中性树胶封固。

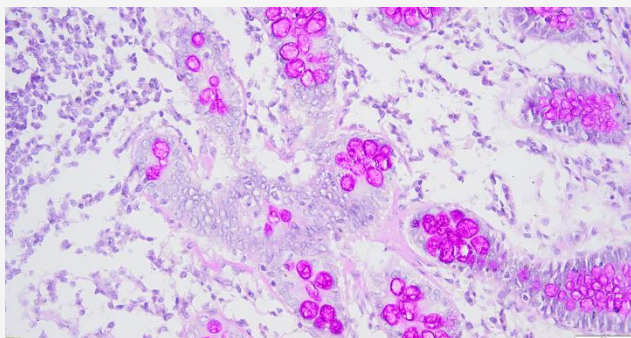
染色结果：

PAS 反应阳性物质(糖原或多糖)	红色或紫红色
细胞核	蓝色
细胞质	深浅不一的红色

备注：颜色深浅很大程度上取决于样品在过碘酸溶液和 Schiff Reagent 中作用时间的长短。

阴性对照(可选)：

- 1、取淀粉酶 1g 溶解于 PBS(pH5.3) 100ml，处理 30 ~ 60min，与其他切片共同入过碘酸溶液。结果应为阴性。
- 2、(备选方案)取唾液片(过滤后用)处理 30 ~ 60min，与其他切片共同入过碘酸溶液。结果应为阴性。
- 3、(备选方案)如果对照片采用其自身样本，对照片不经过碘酸溶液这一步，直接入 Schiff Reagent，结果应为阴性。



注意事项：

- 1、切片脱蜡应尽量干净，否则影响染色效果。
- 2、过碘酸氧化时间不宜过久，氧化时的温度以 18 ~ 22℃最佳。
- 3、过碘酸溶液和 Schiff Reagent 应置于 4℃密闭保存，使用时避免接触过多阳光和空气，使用前，最好提前 30min 取出恢复到室温，避光暗处使用。
- 4、酸性乙醇分化液应经常更换新液，其分化时间应该依据切片厚薄、组织的类别和分化液的新旧而定，另外分化后自来水冲洗时间应该足够。
- 5、在过碘酸溶液和 Schiff Reagent 中作用时间非常重要，该依据切片厚薄、组织的类别等决定。

糖原 PAS 染色液(细胞专用) (DG0006), 糖原 PAS 染色液(真菌专用) (DG0010), 因为其过碘酸溶液和苏木素溶液浓度更低, 不宜过染。

- 7、冷冻切片染色时间尽量要短。
- 8、试剂开封后请尽快使用, 以防影响后续实验效果。

有效期: 12 个月有效。低温运输, 按要求保存。

相关产品:

产品编号	产品名称
CS0001	ACK 红细胞裂解液(ACK Lysis Buffer)
DC0032	Masson 三色染色液
DG0005	糖原 PAS 染色液
DH0006	苏木素伊红(HE)染色液(醇溶)
DJ0001	普鲁士蓝染色液(核固红法)
PE0103	Acr-Bis(30%,29:1)
TE0002	碱性磷酸酶(ALP)检测试剂盒(PNP 微板法)

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